

# **Molybdenum-based polyoxometalates as highly active and selective catalysts for the epimerization of aldoses**

## **Supporting information**

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**Table S1.** The effect of pH on the epimerization of glucose using  $\text{H}_3\text{PMo}_{12}\text{O}_{40}$  as catalyst<sup>a</sup>

Entry	PH	Time (min)	Temperature (°C)	Glucose conversion (%)	Mannose Selectivity (%)
1	1.14	60	80	26.2±1.1	91
2	1.69	60	80	25.6±0.6	90
3	2.37	60	80	25.5±0.9	93
4	3.78	60	80	26.1±0.8	91
5	4.12	60	80	25.4±0.8	94
6	4.86	60	80	22.6±0.7	92
7	5.02	60	80	16.7±0.3	94
8	5.13	60	80	14.7±0.5	95

<sup>a</sup>Reaction condition: For all the reactions, the molar ratio of Mo to glucose was kept constant at 1:50 for all the runs. pH of the solutions was adjusted using concentrated HCl or  $\text{NH}_4\text{OH}$ .

**Table S2.** The effect of pH on the epimerization of glucose using  $\text{Ag}_{3.03}\text{PMo}_{12}\text{O}_{40}$  as catalyst<sup>a</sup>

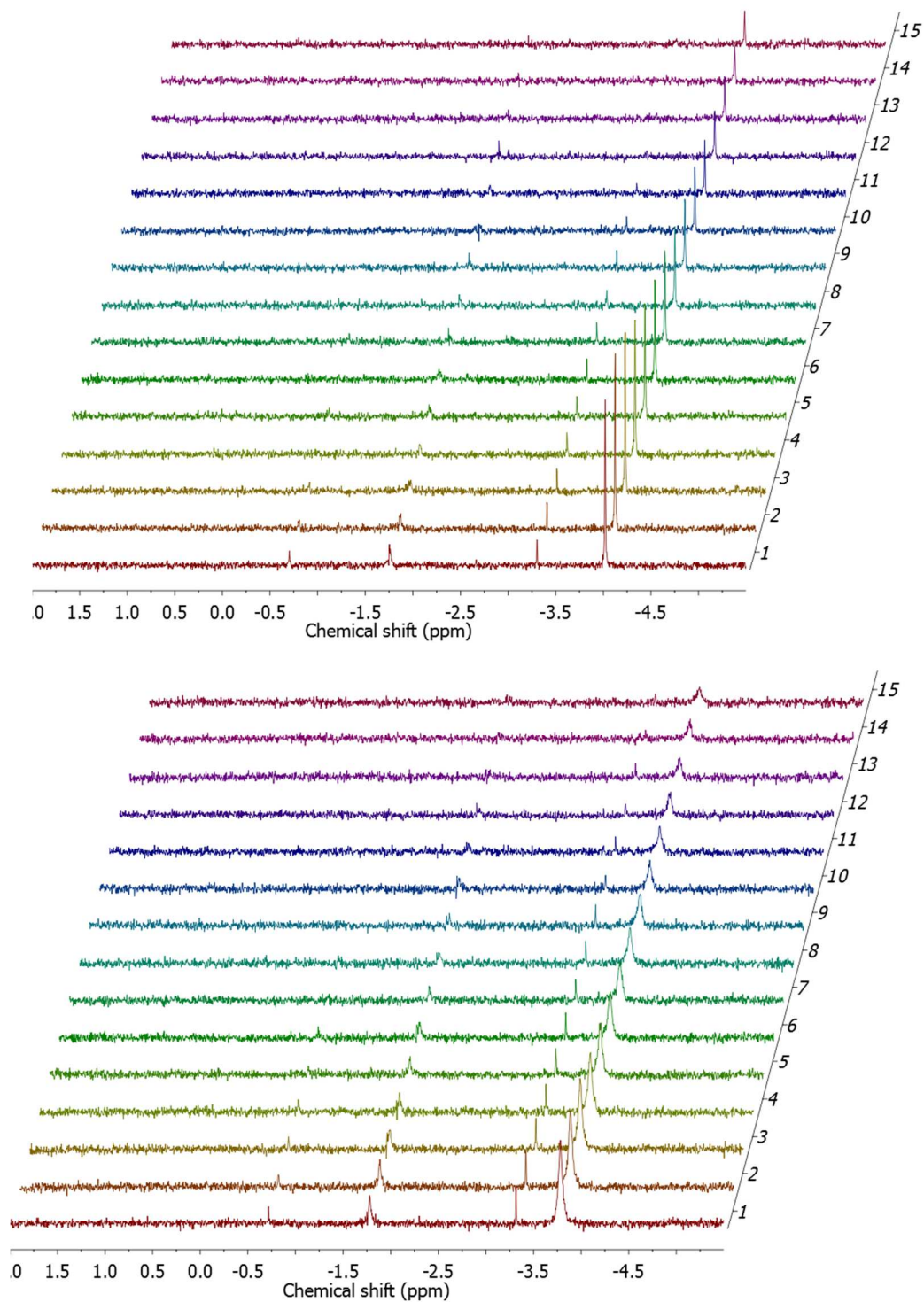
Entry	PH	Time (min)	Temperature (°C)	Glucose conversion (%)	Mannose Selectivity (%)
1	1.22	60	80	25.1±0.8	92
2	1.60	60	80	24.8±0.6	91
3	2.41	60	80	25.3±0.9	90
4	3.68	60	80	26.0±0.8	92
5	4.06	60	80	24.5±0.6	95
6	4.90	60	80	21.7±0.5	93
7	5.01	60	80	15.3±0.5	94
8	5.22	60	80	12.6±0.4	93

<sup>a</sup>Reaction conditions: For all the reactions, the molar ratio of Mo to glucose was kept constant at 1:50 for all the runs. pH of the solutions was adjusted using concentrated HCl or  $\text{NH}_4\text{OH}$ .

**Table S3.** The effect of pH on the epimerization of glucose using  $\text{Sn}_{0.76}\text{PMo}_{12}\text{O}_{40}$  as catalyst<sup>a</sup>

Entry	PH	Time (min)	Temperature (°C)	Glucose conversion (%)	Mannose Selectivity (%)
1	1.18	60	80	25.4±1.0	90
2	1.56	60	80	25.0±1.1	91
3	2.38	60	80	25.7±0.8	94
4	3.75	60	80	25.2±0.7	93
5	4.10	60	80	24.7±0.5	91
6	4.88	60	80	20.8±0.7	92
7	4.97	60	80	14.4±0.4	92
8	5.06	60	80	12.3±0.6	94

<sup>a</sup>Reaction condition: For all the reactions, the molar ratio of Mo to glucose was kept constant at 1:50 for all the runs. pH of the solutions was adjusted using concentrated HCl or  $\text{NH}_4\text{OH}$ .



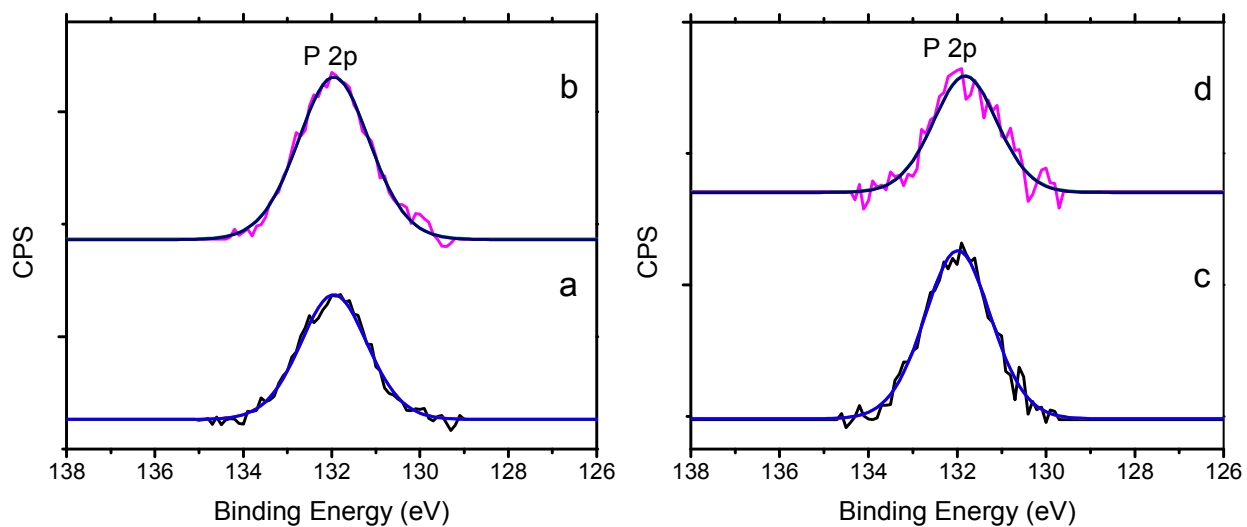
**Figure S1.**  $^{31}\text{P}$  DOSY NMR spectra of 0.1 M of  $\text{H}_3\text{PMo}_{12}\text{O}_{40}$  aqueous solution with glucose (a) before and (b) after reaction

**Table S4.** Chemical shift and width of different species in  $\text{H}_3\text{PMo}_{12}\text{O}_{40}$  solution without glucose

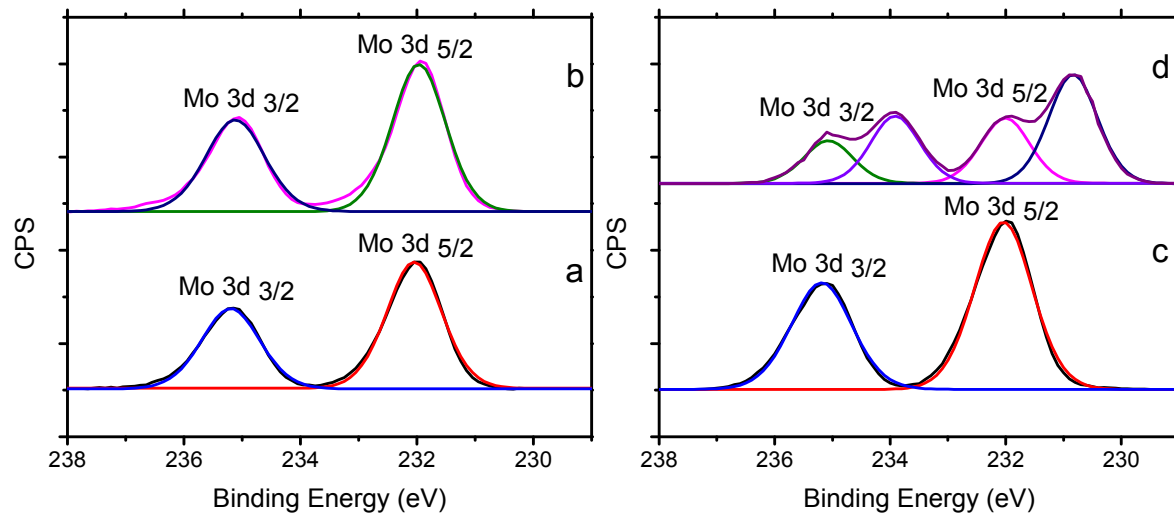
Species	Chemical shift (ppm)		Width (ppm)	
	Before reaction	After reaction	Before reaction	After reaction
1	-0.70	-0.70	3.80	3.50
2	-1.78	-1.78	7.74	8.15
3	-3.29	-3.29	2.91	2.17
4	-4.01	-4.01	2.26	2.30

**Table S5.** Chemical shift and width of different species in  $\text{H}_3\text{PMo}_{12}\text{O}_{40}$  solution with glucose

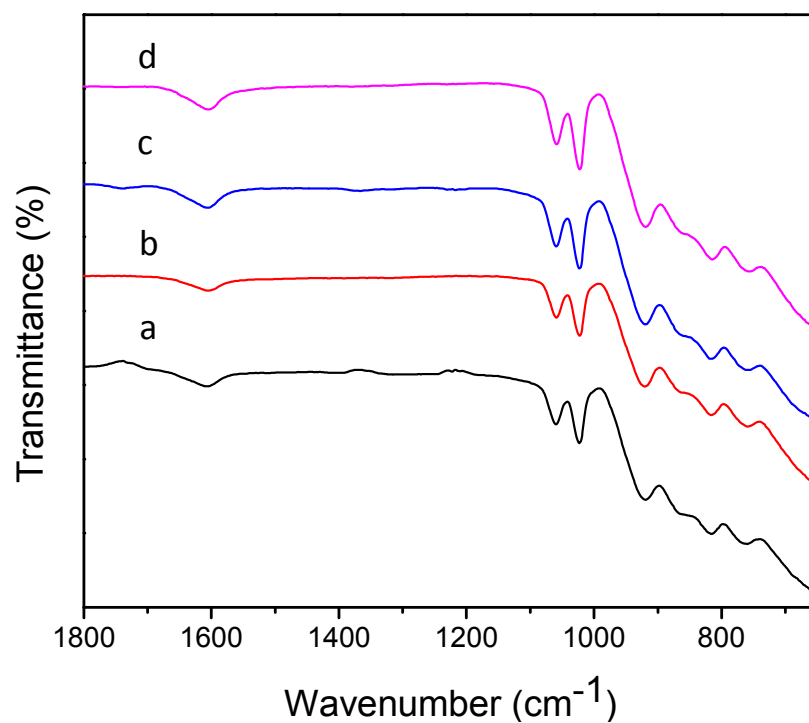
Species	Chemical shift (ppm)		Width (ppm)	
	Before reaction	After reaction	Before reaction	After reaction
1	-0.74	-0.74	3.92	3.74
2	-1.80	-1.80	7.52	6.63
3	-3.34	-3.34	2.18	2.36
4	-4.03	-3.92	3.02	5.70



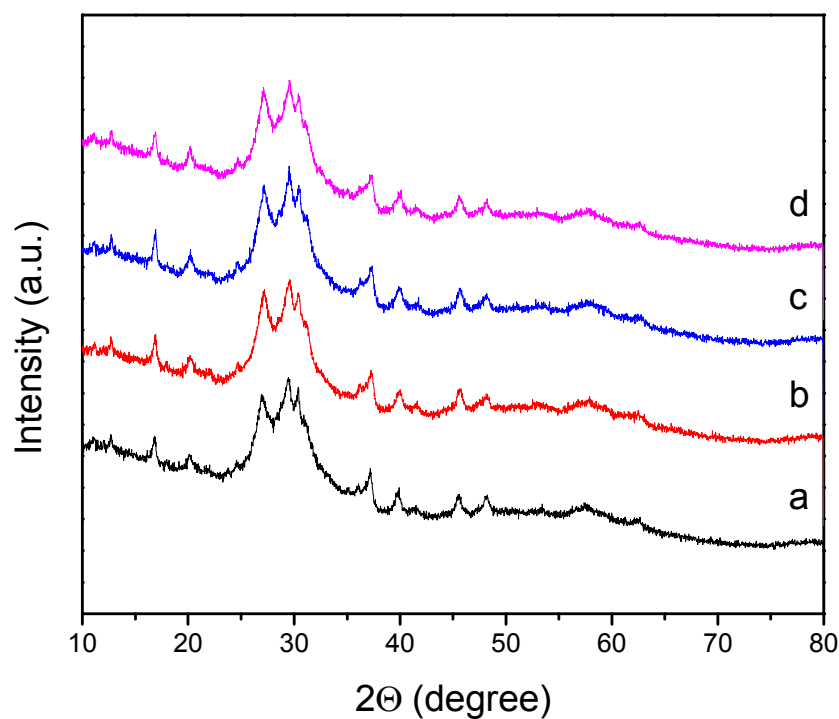
**Figure S2.** XPS spectra of P in  $\text{Ag}_3\text{PMo}_{12}\text{O}_{40}$  precipitated by  $\text{AgNO}_3$  from  $\text{H}_3\text{PMo}_{12}\text{O}_{40}$  solution without glucose (a) before and (b) after reaction; with glucose (c) before and (d) after reaction. (No P oxidation state change was detected.)



**Figure S3.** XPS spectra of Mo in  $\text{Sn}_{0.75}\text{PMo}_{12}\text{O}_{40}$  precipitated by  $\text{SnCl}_4$  from  $\text{H}_3\text{PMo}_{12}\text{O}_{40}$  solution without glucose (a) before and (b) after reaction; with glucose (c) before and (d) after reaction.



**Figure S4.** FTIR spectra of  $\text{Ag}_3\text{PMo}_{12}\text{O}_{40}$  obtained from precipitating  $\text{PMo}_{12}\text{O}_{40}^{3-}$  from solution using  $\text{AgNO}_3$  (a) before and (b) after heating at  $80^\circ\text{C}$  for 60 min in the absence of glucose; (c) before and (d) after reaction with glucose at  $80^\circ\text{C}$  for 60 min.



**Figure S5.** XRD patterns of  $\text{Ag}_3\text{PMo}_{12}\text{O}_{40}$  obtained from precipitating  $\text{PMo}_{12}\text{O}_{40}^{3-}$  from solution using  $\text{AgNO}_3$  (a) before and (b) after heating at  $80^\circ\text{C}$  for 60 minutes in the absence of glucose; (c) before and (d) after reaction with glucose at  $80^\circ\text{C}$  for 60 minutes.



### S.3. <sup>13</sup>C kinetic isotope effect calculation based on experimental data

Figure 8 shows the fits of the experimental data to kinetic models assuming the reversible glucose epimerization is a first-order reaction. The formation of mannose follows the equation:

$$C_m(t) = C_{m,e} \left(1 - e^{-\frac{t}{\tau}}\right)$$

Where  $C_m(t)$  and  $C_{m,e}$  are the concentrations of mannose at time  $t$  and equilibrium, respectively, and  $\tau$  is the time constant for the approach to equilibrium.

Initial turnover rates for unlabeled and labeled glucose were calculated from extrapolation to zero time and non-zero values for rates of formation of mannose.

The <sup>13</sup>C kinetic isotope effect can be calculated as follows:

$$\frac{k(\text{glucose})}{k(13\text{C-glucose})} = \frac{r(\text{glucose})/C(\text{glucose})}{r(13\text{C-glucose})/C(13\text{C-glucose})} \quad \text{where } k \text{ is the rate constant, } r \text{ represents reaction rate and } C \text{ is concentration.}$$

At  $t = 0$  min,  $C(\text{glucose}) = C(13\text{C-glucose})$

$$\text{so } \frac{k(\text{glucose})}{k(13\text{C-glucose})} = \frac{r(\text{glucose})}{r(13\text{C-glucose})} = \frac{\text{Slope}(\text{glucose})}{\text{Slope}(13\text{C-glucose})} = 1.06 \pm 0.03.$$

### S.4. <sup>13</sup>C kinetic isotope effect via theoretical estimation:

The theoretical kinetic isotope effect is estimated via the equation derived from transition state theory.

$$\frac{k(12\text{C-12C})}{k(13\text{C-12C})} = \exp\left(\frac{ZPE(12\text{C-12C}) - ZPE(13\text{C-12C})}{kT}\right) \quad \text{Eq S1}$$

where  $ZPE = \frac{1}{2} h c \nu$ ,  $h$  is the Planck's constant ( $6.63 \times 10^{-34} \text{ m}^2\text{kg/s}$ ),  $c$  is the speed of light ( $2.998 \times 10^8 \text{ m/s}$ ),  $\nu$  is the vibrational frequency of a C-C bond stretching,  $k$  is the Boltzman constant ( $1.38 \times 10^{-23} \text{ m}^2\text{kg/s}^2/\text{K}$ ) and  $T$  is the temperature (353 K).

Substitute  $ZPE$  with  $\frac{1}{2} h c \nu$ , so Eq S1 can be re-written as

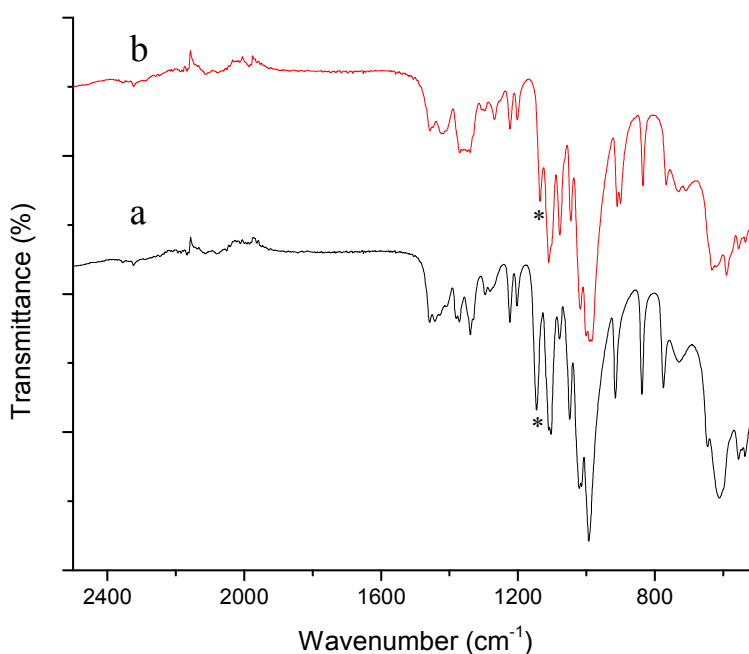
$$\frac{k(12\text{C-12C})}{k(13\text{C-12C})} = \exp\left(\frac{\frac{1}{2} h c (\nu(12\text{C-12C}) - \nu(13\text{C-12C}))}{kT}\right)$$

The stretching vibration between C1 and C2 in unlabeled and <sup>13</sup>C labeled glucose is 1147 and 1136  $\text{cm}^{-1}$ , respectively, based on the FTIR spectra (Figure S6), which has a good agreement

with literature reports.<sup>1-3</sup> So in the equation shown above,  $\nu(12\text{C} - 12\text{C}) = 1147\text{ cm}^{-1}$ ,  $\nu(13\text{C} - 12\text{C}) = 1136\text{ cm}^{-1}$ , and the formation of the C1-C3 bond in mannose is also assumed to proceed via a stretching vibration.

Based on these assumptions, the kinetic isotope effect (KIE), the ratio between the rate constant for epimerization of unlabeled glucose (containing  $^{12}\text{C}1-^{12}\text{C}2$  bond) and the one for epimerization of labeled ( $^{13}\text{C}$ -1)glucose (containing  $^{13}\text{C}1-^{12}\text{C}2$  bond) is  $\frac{k(12\text{C}-12\text{C})}{k(13\text{C}-12\text{C})} = 1.023$

The theoretical KIE is in good agreement with our experimental results shown above.



**Figure S6.** FTIR spectra of (a) unlabeled glucose and (b)  $^{13}\text{C}$  labeled (C1) glucose.

## References

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- (2) Ibrahim, M.; Alaam, M.; El-Haes, H.; Jalbout, A. F.; Leon, A. d. *Eclética Quím.* **2006**, 31, 15-21.
- (3) Urano, T.; Hamaguchi, H. o.; Tasumi, M.; Yamanouchi, K.; Tsuchiya, S.; Gustafson, T. L. *J. Chem. Phys.* **1989**, 91, 3884-3894.